

#### Structure, Measurement & Analysis of Genetic Variation





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### **Learning objectives**

- Get a feel for the variability in the human genome
- What are the most important types of genetic variation?
- Possible functional consequences of genetic variation
- Molecular genetic techniques to individually measure genetic variation (as a prerequisite to investigate its influence on brain imaging traits)

#### Genome variation is visible under the microscope already....



#### ....but it gets enormous at the submicroscopic level

Sequence	<ul> <li>Single nucleotide</li> <li>Base change – substitution – point mutation</li> <li>→ Insertion-deletions ("indels")</li> <li>SNPs – tagSNPs</li> </ul>	Molecular genetic detection
(	<ul> <li>2 bp to 1,000 bp</li> <li>Microsatellites, minisatellites</li> <li>→ Indels</li> <li>Inversions</li> <li>Di-, tri-, tetranucleotide repeats</li> <li>VNTRs</li> </ul>	
Structural variation	<ul> <li>1 kb to submicroscopic</li> <li>→ Copy number variants (CNVs)</li> <li>→ Segmental duplications</li> <li>Inversions, translocations</li> <li>→ CNV regions (CNVRs)</li> <li>Microdeletions, microduplications</li> </ul> Microscopic to subchromosomal <ul> <li>→ Segmental aneusomy</li> <li>Chromosomal deletions – losses</li> <li>Chromosomal insertions – gains</li> <li>Chromosomal inversions</li> <li>Intrachromosomal translocations</li> <li>Chromosomal abnormality</li> <li>→ Heteromorphisms</li> <li>Fragile sites</li> </ul> Whole chromosomal translocations <ul> <li>Interchromosomal translocations</li> <li>Ring chromosomes, isochromosomes</li> <li>Marker chromosomes</li> <li>Aneuploidy</li> </ul>	ſ
	$\rightarrow$ Aneusomy	Cytogenetic

detection

Scherer *et al.,* 2007



#### Single nucleotide polymorphism (SNP)



C-allele: 70% frequency C = major allele

#### **T-allele: 30% frequency**

*T* = *minor* allele

#### Terminology: alleles, genotypes, SNPs,.....



minor allele frequency	inor allele No. requency of SNPs (Mio.)	
1	12.0	290
5	7.1	450
10	5.3	600
20	3.3	960
30	2.0	1,570
40	0.97	3,280

Based on mutation rate and population size it can be assumed that every base pair of the human genome exists in a mutated form in at least several individuals.

- During meiosis, two homologous chromosomes, one from mom and one from dad, twist around each other
- Large segments of DNA are exchanged and recombined
- Gamete (egg cell or sperm cell) is formed, each carries one set of chromosomes from both sides of the parents







#### Recombination does not happen everywhere– The block structure of the human genome



Many SNPs within a block are in "linkage disequilibrium" (LD)



Adapted from Balding DJ (2006) Nature Reviews Genetics.



#### The HapMap Project and 1000Genomes Project define patterns of genetic variation across human genomes





#### An integrated map of genetic variation from 1092 human genomes

The Phase 1 publication, An Integrated map of genetic variation from 1092 human genomes is now available from Nature and can be downloaded directly from the ftp site. The paper is distributed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported licence. Please share our paper appropriately.

All the data files associated with this paper can be found in our phase1 analysis results directory.

LINKS



**All Project** Announcements

#### How many SNPs needed to cover most of the common variation?



# Implications of block structure and linkage disequilibrium for identifying causal alleles for a (imaging) trait



Location of the causative allele (within a gene? where in the gene?) has impact on its functional relevance



Primary transcript

Spliced transcript = mature mRNA, Ready for translation





- 5'-UTR and 3'-UTR important for protein translational control (influence stability of mRNA, subcellular localization, translational control)
- Not part of the protein
- SNPs or mutations in these regions may lead to reduction or acceleration of translation



Primary transcript

- 5'-UTR and 3'-UTR important for protein translational control (influence stability of mRNA, subcellular localization, translational control)
- Not part of the protein
- SNPs or mutations in these regions may lead to reduction or acceleration of translation
- SNPs or mutations in introns may influence gene expression or splicing
- SNPs or mutations in exons may influence protein structure/function (missense, nonsense, frameshift)

#### How to find the SNPs that influence your brain imaging phenotype? Screen the whole genome (GWAS)



#### How to find the SNPs that influence your brain imaging phenotype? Screen the whole genome (GWAS) using SNP arrays



#### **BeadArray Technology (Illumina)**



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### Read-out of the SNP-Arrays



Scan of an individual's DNA with an array harbouring a genome wide set of 550,000 tag SNP markers (Illumina)





#### 5-HTTLPR (serotonin-transporter-linked polymorphic region): a degenerate repeat polymorphism



	<ul> <li>Single nucleotide</li> <li>Base change – substitution – point mutation</li> <li>→ Insertion-deletions ("indels")</li> <li>• SNPs – tagSNPs</li> <li>2 bp to 1,000 bp</li> <li>• Microsatellites, minisatellites</li> <li>→ Indels</li> <li>• Inversions</li> <li>• Di-, tri-, tetranucleotide repeats</li> <li>• VNTRs</li> </ul>	Molecular genetic detection	
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	<ul> <li>Whole chromosomal to whole genome</li> <li>Interchromosomal translocations</li> <li>Ring chromosomes, isochromosomes</li> <li>Marker chromosomes</li> <li>→ Aneuploidy</li> <li>→ Aneusomy</li> </ul>	Cytogenetic	Scherer <i>et al.,</i> 2007

#### **Copy number variants (CNVs) - Definition**

- Variable presence or absence of a DNA sequence >1000 bp
- Different types of CNVs:
  - Deletions
  - Duplications/Triplications
  - Insertions



#### **Copy number variants (CNVs) - Definition**



Wain et al., 2009

#### Larger CNVs (>50-100 kb) can be detected on SNP arrays



#### **Molecular mechansims leading to CNV phenotype**



Lupski & Stankiewicz, 2005

# Most imaging phenotypes will be explained by a spectrum of common and rare functional alleles



New sequencing technologies may help to identify variants relevant for imaging phenotypes not detected so far



# Sequencing whole exomes identifies a lot of "neutral" background variation – how to find the phenotype-relevant variants?

- ~20,000 DNA variants in/near protein coding DNA
- ~200 rare missense variants
- ~100 loss-of-function variants (~20 rare or private)



#### A Systematic Survey of Loss-of-Function Variants in Human Protein-Coding Genes

Daniel G. MacArthur,<sup>1,2\*</sup> Suganthi Balasubramanian,<sup>3,4</sup> Adam Frankish,<sup>1</sup> Ni Huang,<sup>1</sup> James Morris,<sup>1</sup> Klaudia Walter,<sup>1</sup> Luke Jostins,<sup>1</sup> Lukas Habegger,<sup>3,4</sup> Joseph K. Pickrell,<sup>5</sup> Stephen B. Montgomery,<sup>6,7</sup> Cornelis A. Albers,<sup>1,8</sup> Zhengdong D. Zhang,<sup>9</sup> Donald F. Conrad,<sup>10</sup> Gerton Lunter,<sup>11</sup> Hancheng Zheng,<sup>12</sup> Qasim Ayub,<sup>1</sup> Mark A. DePristo,<sup>13</sup> Eric Banks,<sup>13</sup>

A few things are a bit more interpretable (obvious functionality), but not absolute proof in each case...

- ~1 *de novo* variant per exome (only ~5% LoF)
- <5% chance that an individual has a complete knockout of a single well-preserved\* gene anywhere in the genome

\* well-preserved = 98-99% of genes without a common LoF mutation

#### **Functional annotation for variants is crucial:**

influence on methylation of DNA (methylation quantitative trait loci – mQTL)



#### **Summary**

- The genome is highly variable.
- Especially common SNP variants as well as large structural variants (CNVs) can be tested using array-based technologies.
- The field is moving to whole-genome sequencing which allows also detection of rare SNPs and small CNVs/InDels
- Functional annotation of identified genetic variants that might play a role in brain phenotypes is of great importance:

influence on gene regulation (incl. methylation), protein function