

DIAGNOSING FEVER IN RETURNING TRAVELERS

-RECOMMENDATIONS FROM THE BEDSIDE AND LABORATORY

**CARLOS DEL RIO, M.D., MCAP, FIDSA
D. JANE HATA, PH.D., D(ABMM)**

LEARNING OBJECTIVES

- Three illustrative cases of fever in travelers
- Standard of care tests
- Esoteric tests
- New diagnostic approaches
 - Syndromic NAAT panels
- Laboratory workflow

1

CASE DESCRIPTION

CASE # 1

- A 52 y/o women originally from Nigeria returned from a one-month trip in which she visited family and friends that live predominantly in the Borno State in the North-East region of the country.
- She did not take malaria prophylaxis as she grew up in Nigeria and “there was never a need to take medicines” .
- States that as she was getting on the plane to return to the U.S., she started to have chills and later had fevers.
- Since then, she notes having daily evening fevers to 39 – 40°C. She feels tired and has a bad headache.
- Went to a local clinic and was prescribed Acetaminophen which she has taken without improvement.
- On physical exam she is afebrile, her HR is 110 and her BP is 140/70. On exam she is pale, in mild distress and has a palpable spleen tip.
- Laboratory findings include mild anemia and elevated liver enzymes.

TRAVEL HISTORY IS KEY!

- Estimated that 8% of travelers require medical care
 - 30% require hospitalization
- When did travel occur?
 - Variable incubation periods
- Type of travel and duration
 - Urban?
 - Rural?
 - Ecotourism/Adventure tourism?
 - Other exposures (animals, plants, insects, food, water)?
- Sexual exposures
- Refugee/Migrant status
- Onset and nature of symptoms
 - Duration of fever
 - Other symptoms



Sondén K et al. Travel Med Infect Dis. 2025 May-Jun;65:102861.

Thwaites GE, Day NP. N Engl J Med. 2017 Feb 9;376(6):548-560.

INITIAL LABORATORY TESTING

- CBC and differential
- Liver enzymes
- Blood cultures
- Blood smear for malaria and/or rapid antigen diagnostic test (RDT)
 - RDT must be confirmed by smear
- Lumbar puncture and CSF analysis
- Other imaging
- Patient presentation and history will drive additional testing
 - Diarrhea, respiratory symptoms, rash, jaundice

Thwaites GE, Day NP. N Engl J Med. 2017 Feb 9;376(6):548-560.
Wu HM. Semin Diagn Pathol. 2019 May;36(3):197-202.

“THINK HORSES, NOT ZEBRAS”

-Dr. Theordore Woodward, Univ of Maryland

MOST LIKELY PATHOGENS; “THE HORSES”

Most Common

- Malaria
 - Species related to geography
- Dengue Fever
- EBV
 - Mononucleosis
- Rickettsial infections
 - Typhus, RMSF, ehrlichiosis, *Anaplasma*
- *Salmonella enterica*
 - Serotype typhi, paratyphi
 - Enteric fever
- “Other viral infections”

Also Consider

- Influenza
- Sars CoV-2
- ETEC
- Zika
- Chikungunya
- MDR bacteria
- TB
- Regional outbreaks

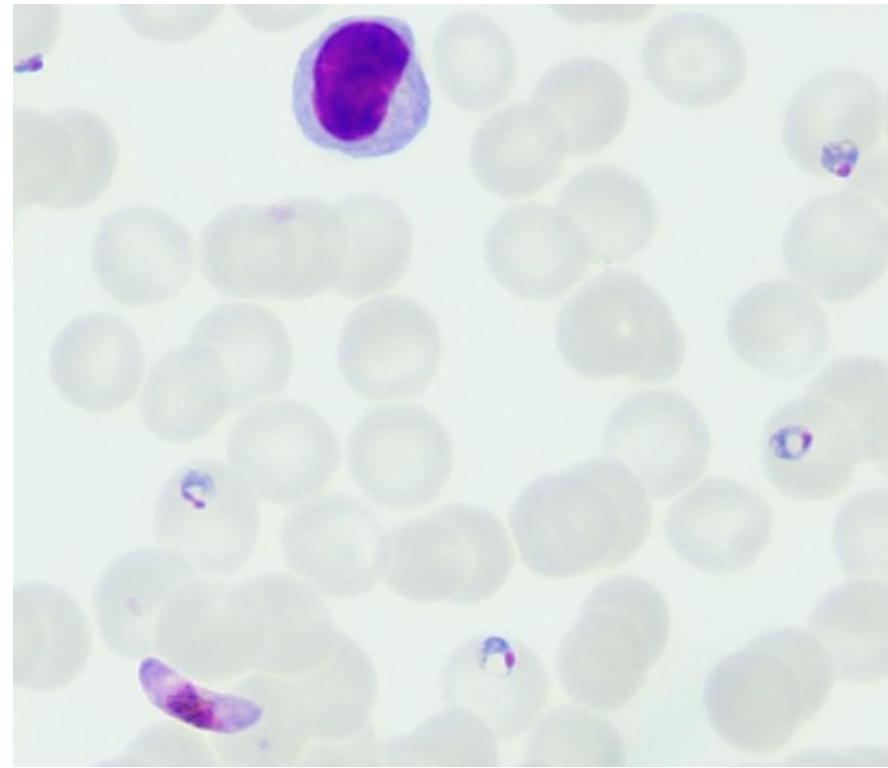


<https://www.uptodate.com/contents/evaluation-of-fever-in-the-returning-traveler>

Brown AB, et al. MMWR Surveill Summ. 2023 Jun 30;72(7):1-22.

CASE #1 - MALARIA

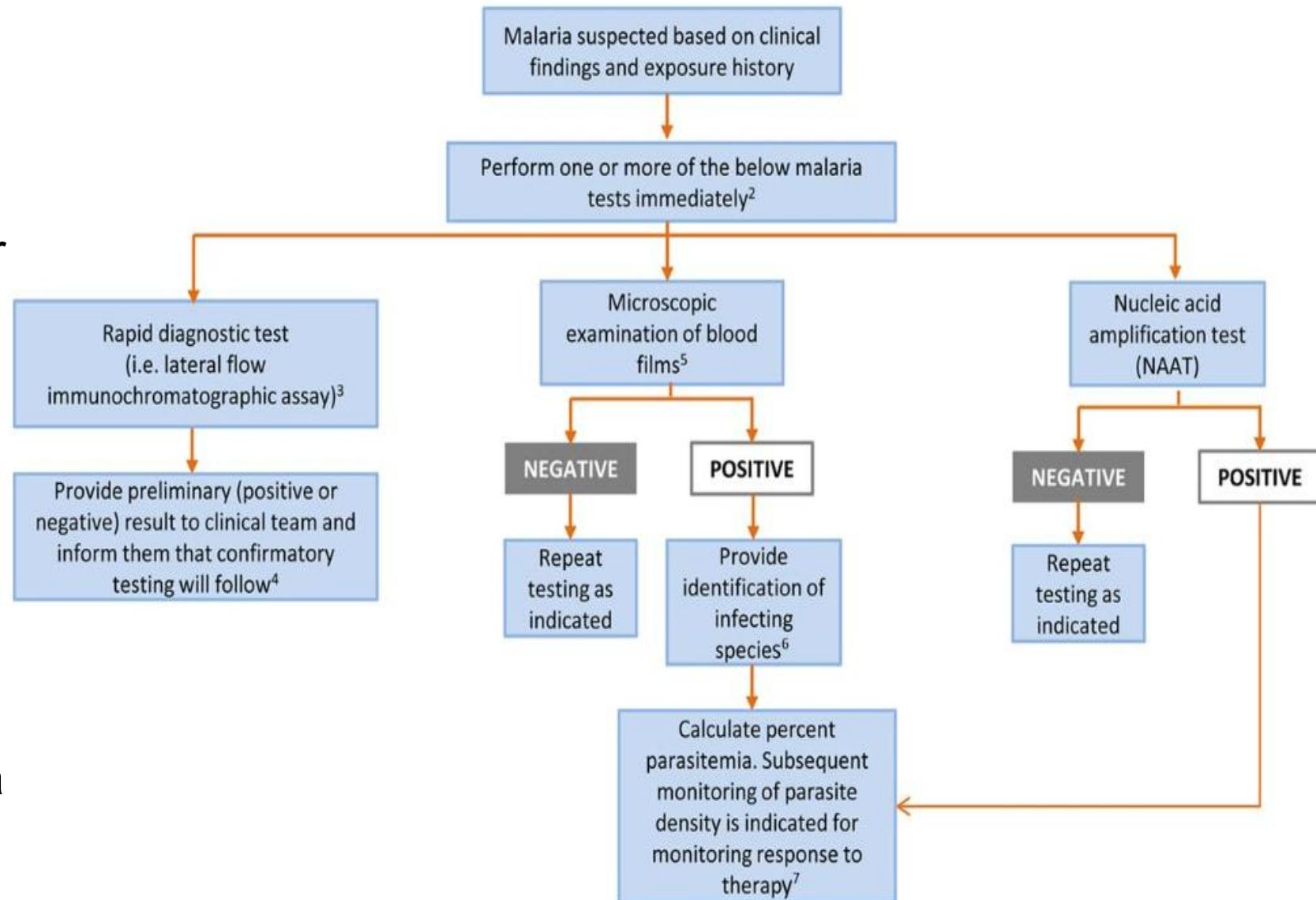
- Most likely cause of fever in travelers
 - *P. falciparum, P. vivax, P. ovale, P. malariae*
- Laboratory should have ability to screen STAT
 - Thick and thin smears
 - Detection of other pathogens (*Babesia, Ehrlichia, Trypanosomes*)
- Speciation from thin smears
 - Calculation of percent parasitemia
- Rapid antigen tests
 - Lateral flow immunochromographic method
 - 1 test FDA approved for use
 - Low sensitivity at ≤ 200 parasites/microliter
- NAATs
 - Not widely available, may not be STAT test



Christmas Eve 2024

LABORATORY ALGORITHM FOR MALARIA

- Smears still considered gold standard
 - Repeat every 6-8 hours for up to 3 days
- Speciation critical for clinical management
- Quantification
 - Severe malaria $\geq 5\%$ parasitemia
- RDT requires smear confirmation
- NAATs require smear confirmation for % parasitemia



LESS LIKELY PATHOGENS; “THE ZEBRAS”

- Viral infections
 - Measles
 - Viral hemorrhagic fevers
 - Ebola
 - Marburg
 - Rabies
 - WNV
 - Mpox
 - Oropouche
- Parasitic infections
 - Leishmaniasis
 - Various helminths
 - *Entamoeba histolytica*
- Bacterial infections
 - Brucellosis
 - Tetanus
 - Melioidosis
 - Q Fever
- US Travel
 - Plague
 - Babesiosis
 - *Coccidioides immitis/posadasii*
 - Ehrlichiosis/Anaplasmosis
 - Hantavirus
- Emerging pathogens



<https://www.uptodate.com/contents/evaluation-of-fever-in-the-returning-traveler>

Brown AB, et al. MMWR Surveill Summ. 2023 Jun 30;72(7):1-22.

TYPES OF TESTS AVAILABLE

-WHAT CAN YOUR LAB DO?



Culture/Microscopy

- Bacterial ID
 - Phenotypic
 - MALDI-TOF
- AST methods
- Ova and Parasite exam
- Malaria smears

Serology

- Antibody detection
- Antigen detection
- Immuno-chromatographic
 - Lateral-flow devices
- Immunoassay
 - Automation

Molecular

- NAATs
 - LDTs
- Single analyte
- Multiplex
- Syndromic panels
- NGS

SYNDROMIC TESTING NAAT PANELS POST-TRAVEL

-ARE THESE USEFUL?

Respiratory

- Adenovirus
- Coronavirus 229E, HKU1, NL63, OC43
- SARS-CoV-2
- HMPV
- Human Rhinovirus/Enterovirus
- Influenza A virus
- Influenza A virus A/H1
- Influenza A virus A/H3
- Influenza A virus A/H1-2009
- Influenza B virus
- RSV

- *Bordetella parapertussis*
- *Bordetella pertussis*
- *Chlamydia pneumoniae*
- *Mycoplasma pneumoniae*

Gastrointestinal

- Astrovirus
- Adenovirus F40/41
- Norovirus GI/GII
- Rotavirus A
- Sapovirus (I, II, IV, and V)

- *Campylobacter* (*C. jejuni* / *C. coli* / *C. upsaliensis*)
- *Clostridioides* (*Clostridium*) *difficile* (toxin A/B)
- *Plesiomonas shigelloides*
- *Salmonella*
- *Yersinia enterocolitica*
- *Vibrio* (*V. parahaemolyticus* / *V. vulnificus* / *V. cholerae*)
- EAEC, EPEC, ETEC, STEC EIEC

- *Cryptosporidium*
- *Cyclospora cayetanensis*
- *Entamoeba histolytica*
- *Giardia lamblia*

Meningitis/Encephalitis

- CMV
- Enterovirus
- HSV 1,2
- HHV-6
- Human parechovirus
- VZV

- *Escherichia coli* K1
- *Haemophilus influenzae*
- *Listeria monocytogenes*
- *Neisseria meningitidis*
- *Streptococcus agalactiae*
- *Streptococcus pneumoniae*

- *Cryptococcus* (*C. neoformans*/*C. gattii*)

SYNDROMIC TESTING NAAT PANELS POST-TRAVEL

-ARE THESE USEFUL?

Respiratory

- Adenovirus
- Coronavirus 229E, HKU1, NL63, OC43
- SARS-CoV-2
- HMPV
- Human Rhinovirus/Enterovirus
- Influenza A virus
- Influenza A virus A/H1
- Influenza A virus A/H3
- Influenza A virus A/H1-2009
- Influenza B virus
- RSV

- *Bordetella parapertussis*
- *Bordetella pertussis*
- *Chlamydia pneumoniae*
- *Mycoplasma pneumoniae*

Gastrointestinal

- Astrovirus
- Adenovirus F40/41
- Norovirus GI/GII
- Rotavirus A
- Sapovirus (I, II, IV, and V)

- *Campylobacter* (*C. jejuni* / *C. coli* / *C. upsaliensis*)
- *Clostridioides* (*Clostridium*) *difficile* (toxin A/B)
- *Plesiomonas shigelloides*
- *Salmonella*
- *Yersinia enterocolitica*
- *Vibrio* (*V. parahaemolyticus* / *V. vulnificus* / *V. cholerae*)
- EAEC, EPEC, ETEC, STEC EIEC

- *Cryptosporidium*
- *Cyclospora cayetanensis*
- *Entamoeba histolytica*
- *Giardia lamblia*

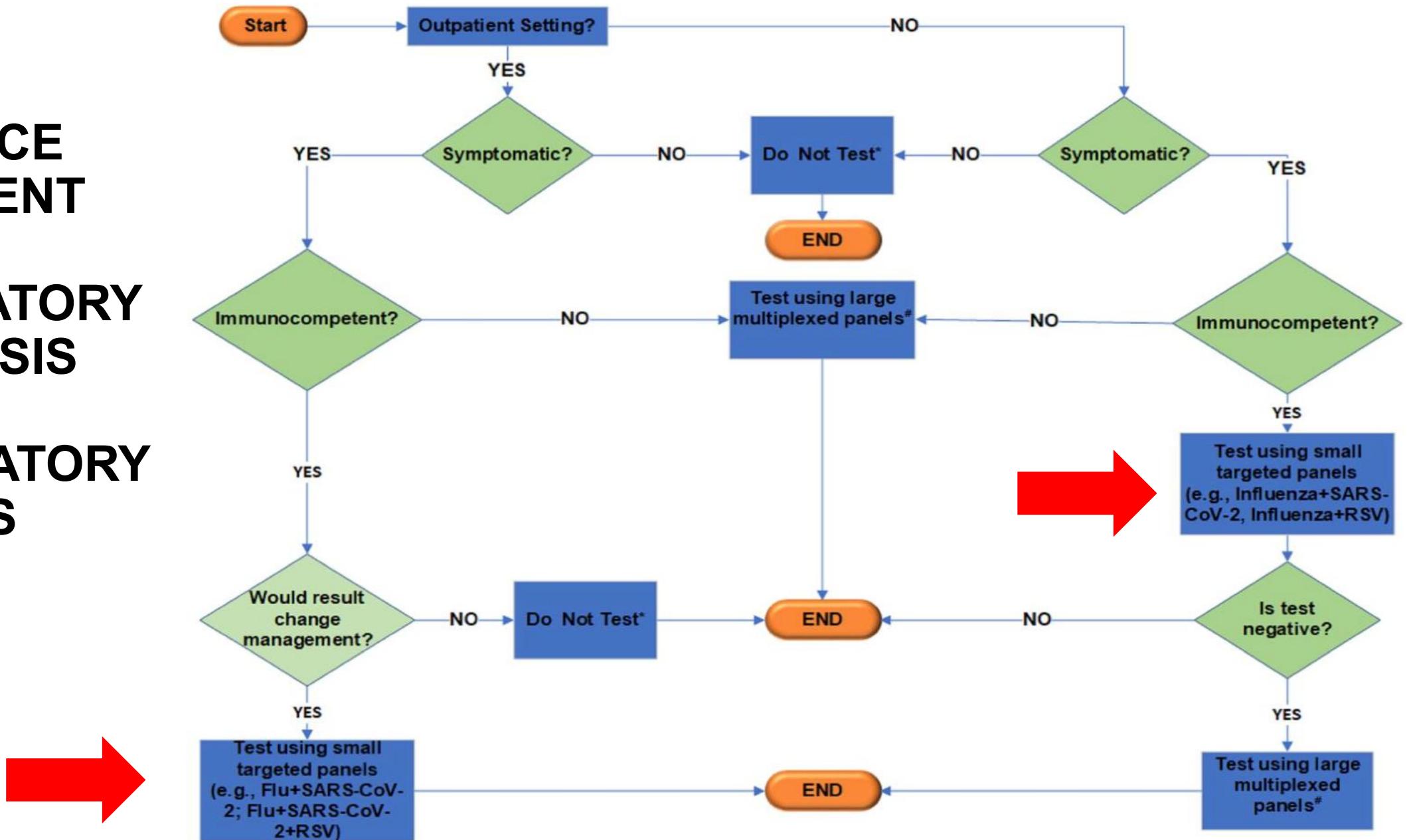
Meningitis/Encephalitis

- Targets not among most likely pathogens
- Use based on clinical presentation

*Bacterial culture still needed for AST, submission to PH

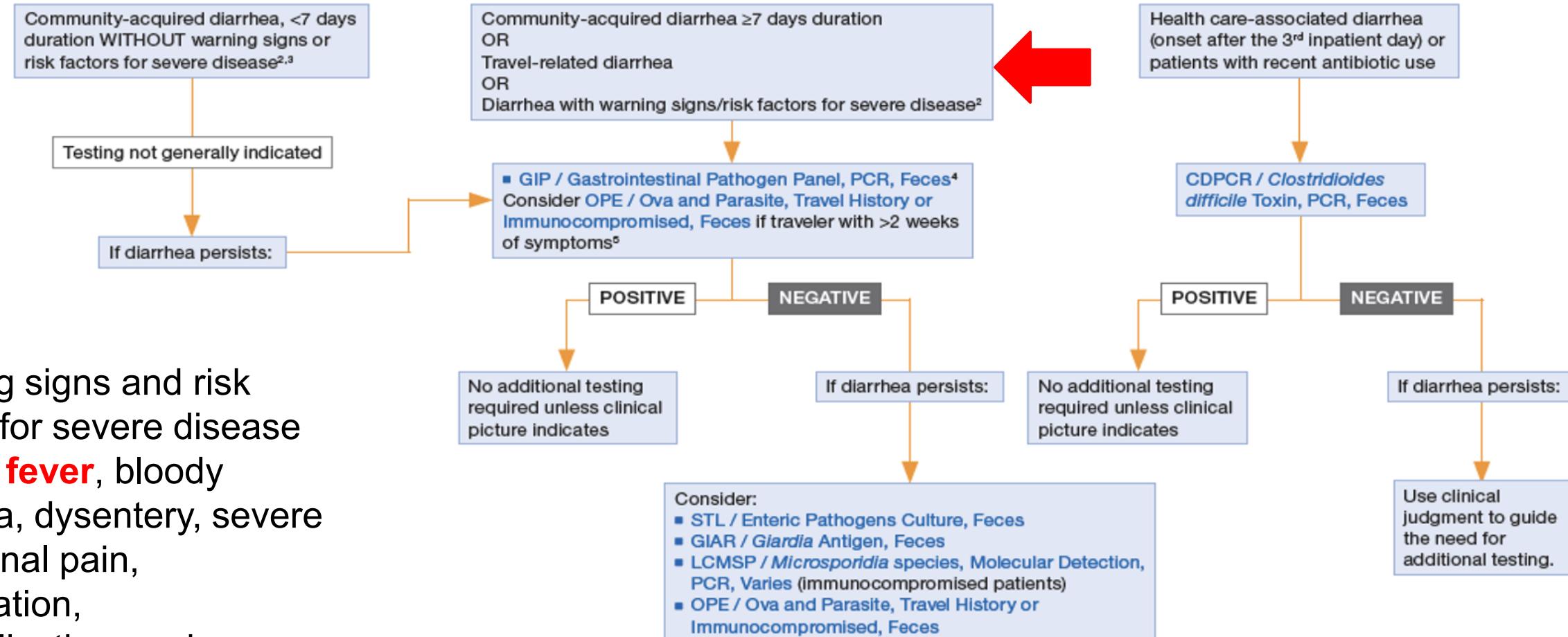
- Consider use of smaller, targeted panels or single NAAT assays

ADLM GUIDANCE DOCUMENT ON LABORATORY DIAGNOSIS OF RESPIRATORY VIRUSES



LABORATORY TESTING FOR INFECTIOUS CAUSES OF DIARRHEA

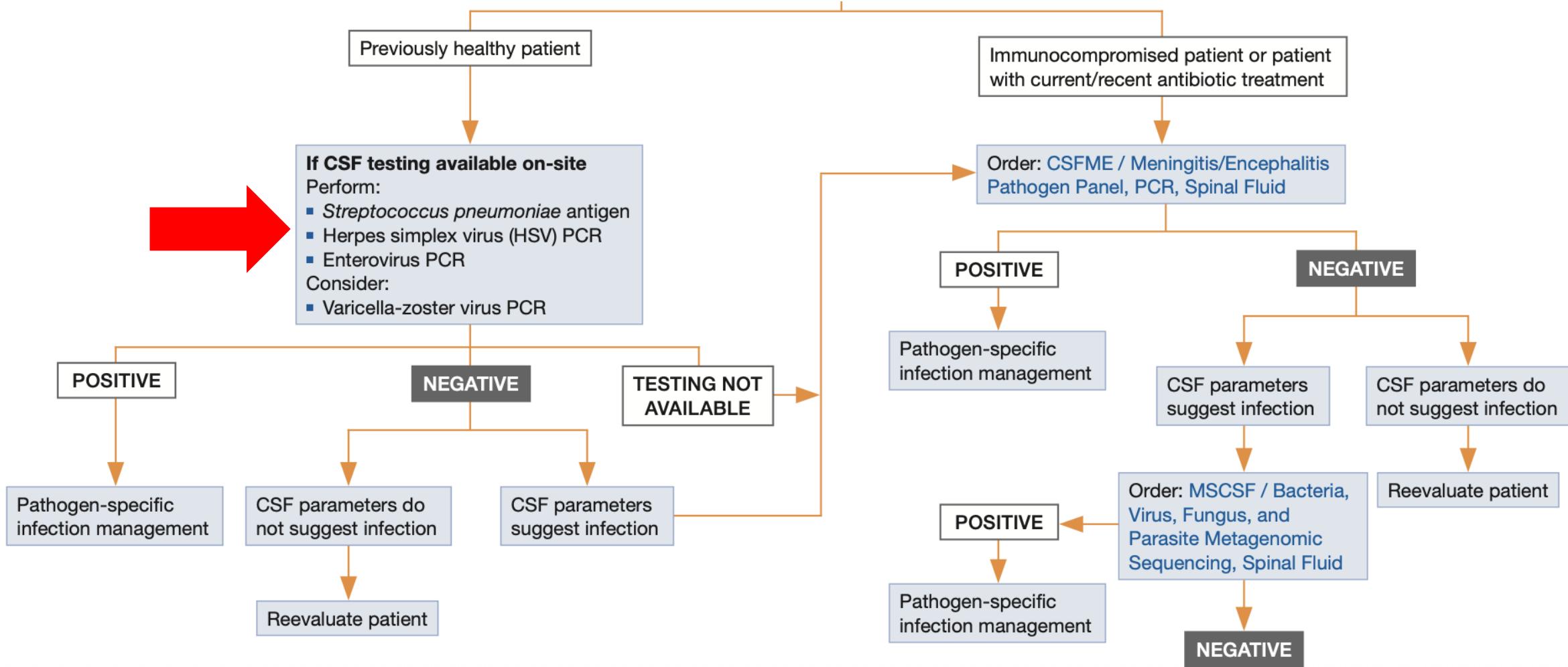
MAYO CLINIC LABORATORIES



Warning signs and risk factors for severe disease include **fever**, bloody diarrhea, dysentery, severe abdominal pain, dehydration, hospitalization, and immunocompromise

MENINGITIS/ENCEPHALITIS ALGORITHM

MAYO CLINIC LABORATORIES



2

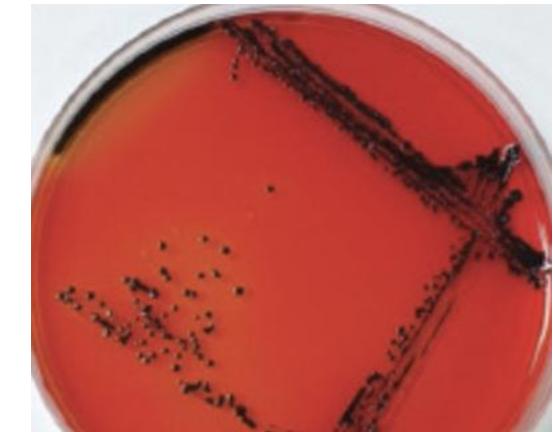
CASE DESCRIPTION

CASE # 2

- A 37 y/o male returned from a 2-week trip to Colombia where he went hiking and backpacking in the jungle.
- He states that during the trip he had a short episode of loose stools.
- Two days after retuning home to the U.S. he developed fever, headache and malaise.
- He has no appetite and feels unwell.
- On physical exam he is febrile to 39° C, his HR is 60 and his BP is 100/60. His exam is otherwise normal with no hepatosplenomegaly.
- Laboratory finding include pancytopenia and elevated liver enzymes.

CASE #2 LABORATORY WORKUP

- Broad differential
- Stool plated to BA, MAC, XLD
- Gastrointestinal NAAT panel performed
 - *Salmonella* sp.
- Gram-negative bacilli on XLD after 2 days of incubation at 37° C.
 - XLD Red colonies with black center (H_2S)
 - MAC no color
- *Salmonella* sp. identified by MALDI-TOF
- Reported to public health laboratory
 - Isolate submitted for serotyping



Salman HA et al. PMCID: PMC8404135

CASE #2

WHAT'S A LAB TO DO?



Culture

- Selective or chromogenic media
 - MAC, S/S, HE, XLD, bismuth sulfate, enrichment broth
- TSI: K/A; H₂S
- Consider turnaround time



Serotyping

- Capsular antigen (Vi)
- Somatic antigen (O)
 - Serogroup D
- > 2500 serotypes
- Generally performed at PH laboratories



Antimicrobial Susceptibility Testing

- Only for invasive and typhoidal isolates
- See CLSI M-100
- AM, CIP, LVX, SXT, CRO, CTX, AZM
- XDR emerging (Middle East, SE Asia)

CASE #2

WHAT'S A LAB TO DO?



MALDI-TOF

- Need pure culture isolate
- ID as *Salmonella* sp. (genus level)
- Database dependent



Molecular

- Multiplex GI NAAT performed direct from specimen
- *Salmonella* sp. (*enterica* and *bongori*)
- Subtypes Typhi and Paratyphi included
- NGS for strain typing



Reporting

- Reportable to Public Health
 - PulseNet
- Organism needed for PH submittal

CASE RESOLUTION

- Identified by public health laboratory by WGS as
 - *Salmonella enterica* subsp. *enterica* serotype Typhi
 - O antigen Serogroup D
- *Salmonella enterica*
 - Six subtypes (I – VI)
- AST results (primary agents): All susceptible
 - Ampicillin
 - Ciprofloxacin/Levofloxacin
 - TMP/SFX
 - Cefotaxime/ceftriaxone
- Emergence of XDR strains with resistance to all first-line agents
 - Pakistan/Iraq; consider carbapenem or azithromycin

3

CASE DESCRIPTION

CASE # 3

- A 5 y/o boy travelled to Italy with his parents to visit friends that live near Milan. They stayed for 5 days.
- Three days after returning the parents noticed that he had a runny nose and a cough. He developed high fever the next day as well as bilateral conjunctivitis.
- He felt unwell and had no appetite.
- Approximately two days later the mother noticed a rash that was mostly in the hairline and behind the ears.
- On physical exam he is febrile to 38.8° C and tachycardic. He has bilateral conjunctivitis and a maculopapular rash in the face and upper body.
- Laboratory finding include leukopenia with lymphopenia and elevated liver enzymes.
- Mother states that she has her family do not believe in vaccines.

CASE #3

-WHAT'S A LAB TO DO?

- Infectious differential (just a few)

Viruses causing rash	Bacteria causing rash
EBV, VZV	<i>Neisseria meningitidis</i>
Primary HIV	Staphylococcal toxic shock
Parvovirus B19	Rickettsial infections
Enterovirus	Typhoid fever

- NP swab submitted for culture and NAAT
 - All analytes negative by respiratory panel
 - Inoculate blood and respiratory cultures
- Clinical presentation can aid diagnosis and guide isolation

Wu HM. Semin Diagn Pathol. 2019 May;36(3):197-202.

MORE INFORMATION COMES TO THE LAB....

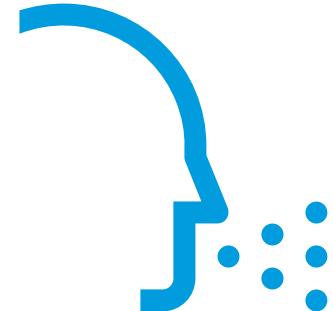


- Patient presentation:
 - Had conjunctivitis, cough, and coryza for 7 days
 - Maculopapular rash on the upper body
 - Unable to confirm vaccination history
- Clinical presentation and history suggests.....

Measles

OPTIONS FOR MEASLES (RUBEOLA) TESTING

- Reportable disease – communicate with your PH lab first for assistance
 - 90% infection rate in susceptible populations
- Identify – Isolate – Inform
 - Handle specimens carefully!
- RT-PCR
 - NP swab, throat swab within 5 days of rash onset
 - Urine within 6-14 days of rash onset
 - Available from public health laboratories, some commercial reference laboratories
 - Laboratory-developed tests could be an option
 - No commercial FDA-approved test available
 - **Measles not included on syndromic NAAT panels**
- Serologic detection of measles IgM
 - Useful for confirmation
 - Negative in early disease
 - Available from public health laboratories, some commercial reference laboratories



ESOTERIC TESTING FOR TRAVEL ASSOCIATED PATHOGENS

“TROPICAL FEVER” SYNDROMIC PANELS

BIOFIRE® FILMARRAY® TROPICAL FEVER (TF) PANEL

- *Leptospira* sp. (Group 1)
- Chikungunya
- Dengue
 - Serotypes 1,2,3,4 grouped together
- *Plasmodium* sp.
 - *P. falciparum*, *P. vivax*, *P. ovale*, *P. malariae*, and *P. knowlesi*
- *Plasmodium falciparum*
- *Plasmodium vivax/ovale*
- Specimen: 200 microliters whole blood (EDTA)



TF ASSAY PERFORMANCE

- From IFU: “Evaluation for more common causes of febrile illness should be considered prior to evaluation with this panel”
- Qualitative results only
 - Quantitation of *Plasmodium* sp. requires smear
- PCR may over diagnose disease in patients from endemic areas

Target	N (pos/neg)	PPA%	NPA%
<i>Leptospira</i> sp.	16/1859	93.8	99.8
Chikungunya	25/1850	100	99.9
Dengue (1-4)	283/1592	94.0	100
<i>Plasmodium</i> sp.	345/1530	98.3	99.2
<i>P. falciparum</i>	248/1627	92.7	99.8
<i>P. vivax/ovale</i>	124/1751	92.7	99.8

OTHER ESOTERIC TESTS

HOW TO GET THESE DONE?

- Consider turnaround time and need for clinical intervention
- Serology
 - State PH labs/CDC – check test availability
 - Commercial reference labs
- Other NAAT testing
 - State PH labs/CDC – check test availability
 - Commercial reference labs
- Role of NGS
 - Availability
 - Turnaround time
- Viral culture
 - Few performing laboratories; potentially hazardous
- Develop an LDT?

Consider what
you will do before
it happens

LABORATORY COMMITMENT IF ESOTERIC TESTING PERFORMED

Purchase of test platform

Test verification/ validation

Training and competency of staff

Proficiency testing

Quality control/quality assurance

Reagent inventory management

Laboratory workflow

- Random access?
- Once per day?
- Once per week?
- What shift?

GENERAL TESTING COMMENTS

LABORATORY BIOSAFETY

- May evolve based on identification of pathogen
- Standard PPE
 - Impermeable gown/lab coat
 - Gloves
 - Additional respiratory protection depending on pathogen
- Use of BSC for specimen manipulation
 - Inoculation of culture plates
 - Creating smears
 - NAAT inoculation
- Use of automated test systems
 - Follow manufacturer recommendations for cleaning and disinfection
- Biohazardous waste disposal



LABORATORY WORKFLOW

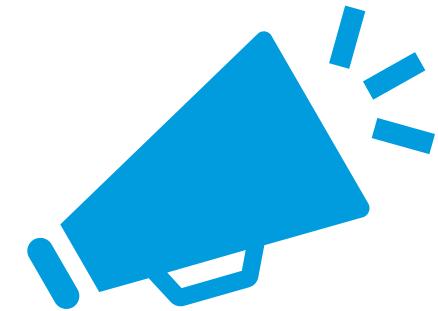
- Consider scenarios ahead of time
 - *What might you be likely to see?*
 - Institution and laboratory drills
- Develop plans through collaboration
 - Infectious Disease
 - Infection Control
 - Clinical services (ED, outpatient)
- Consider what testing is feasible for your laboratory practice
 - Academic or referral center?
 - Small hospital or community hospital?
 - Nature of patient population



HCID Drill June 2025

COMMUNICATION OF FINDINGS

- Critical/alert value?
- Infection control considerations
- Notification of Public Health
- Key Groups
 - Infectious Disease
 - Infection Prevention
 - Who are your contacts?
- High Consequence Infectious Disease plan?
- Plan for laboratory handling of biological agents?



RESOURCES

- State PH laboratories and CDC
- CDC Yellow Book
 - [https://www.cdc.gov/travel/yellow-book/index.html](https://www.cdc.gov/travel/yellowbook/2020/introduction/introduction-to-the-yellow-book)
- WHO
 - <https://www.who.int/emergencies/diseases-outbreak-news>
- GeoSentinel
 - https://geosentinel.org/alerts/?post_type=alert&s=2025
 - Membership required
- ProMED
 - <https://www.promedmail.org>
 - Membership required



THANK YOU AND SAFE TRAVELS!